

Antiviral Characterization of a Next Generation Chemical Series of HBV Capsid Inhibitors *In Vitro* and *In Vivo*

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BACKGROUND

- Hepatitis B virus (HBV) replication is strictly dependent upon capsid assembly around pregenomic RNA (pgRNA)
- Proper assembly of HBV nucleocapsid is essential for viral genome relaxed circular DNA (rcDNA) synthesis, infectious virion production and maintenance of a nuclear covalently closed circular DNA (cccDNA) pool
- The capsid assembly process thus represents a *bona fide* antiviral target, and constitutes a novel mechanism that is distinct from the nucleos(t)ide analogues currently available for clinical use
- Interfering with HBV capsid assembly with small molecule inhibitors has been shown to translate into antiviral activity *in vitro* and *in vivo* (Cole, 2016)

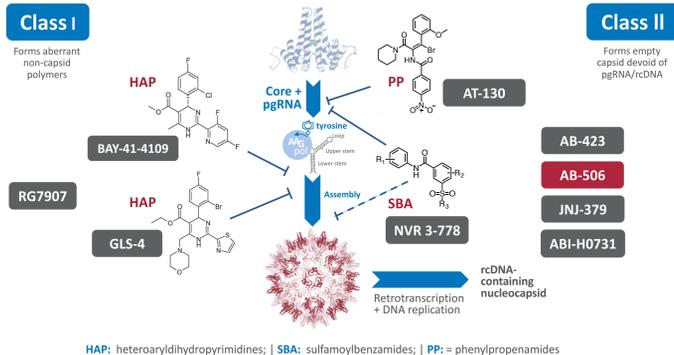


Figure 1: HBV capsid assembly pathway and examples of capsid inhibitors.

OBJECTIVES

Characterize the *in vitro* and *in vivo* antiviral activities of a next generation chemical series of potent small-molecule inhibitors of HBV capsid assembly.

MATERIALS AND METHODS

- Compounds were tested in a biochemical assay of capsid assembly as described previously (Zlotnick, 2007)
- The ability of compounds to bind and thermally stabilize core protein was determined in thermal shift assays using differential scanning fluorimetry
- X-ray crystallography studies were conducted to determine the binding mode of compounds to core protein Cp-Y132A mutant
- Antiviral activity was determined in different cell culture models of HBV using branched DNA, quantitative PCR, and AlphaLISA® assays to measure effects on rcDNA or secreted e-antigen
- Activity against HBV genotypes and nucleoside analog inhibitor-resistant (Nuc^R) variants was determined using a transient transfection assay system
- Cytotoxicity of compounds was evaluated in various cell lines using CellTiter-Glo® or MTT assay
- Antiviral activity against viruses of various families was determined using cell culture assays
- Pharmacokinetic profiles of the compound were determined in CD-1 mice, SD rats, and Beagle dogs
- The *in vivo* antiviral activity was assessed in a hydrodynamic injection (HDI) HBV mouse model utilizing pHBV1.3 (Guidotti 1995). Test articles were administered orally for 7 days starting on Day 0, AB-506 and vehicle twice daily and ETV once daily. HBV DNA was measured using qPCR. Reported liver HBV DNA values are vector-subtracted

RESULTS

Table 1: *In vitro* antiviral activities of next generation capsid inhibitors:

Compound	HepDE19 (rcDNA_bDNA) (μM)			HepBHAe82 (HBeAg AlphaLISA) (μM)			HepG 2.2.15 (HBV DNA qPCR) (μM)	
	EC ₅₀	EC ₉₀	CC ₅₀	EC ₅₀	EC ₉₀	CC ₅₀	EC ₅₀	CC ₅₀
Compound A	0.11	0.45	>25	0.07	0.28	>25	0.08	>10
Compound B	0.06	0.27	>25	0.03	0.14	>25	0.04	>10
Compound C	0.04	0.14	15	0.01	0.07	22	0.04	>10
AB-506	0.07	0.27	>25	0.04	0.20	>25	0.04	>10

Table 2: Antiviral activity against HBV genotypes A through D and potency of AB-506 against Nuc^R variants:

HBV Nuc ^R Variant	HBV DNA qPCR		HBV Genotype	HBV DNA qPCR	
	AB-506 (EC ₅₀ μM)			AB-506 (EC ₅₀ μM)	
rtM204I	0.059		A1	0.008	
rtM204I + V173L	0.038		A2	0.023	
rtM204I + S202G	0.052		B1	0.017	
rtM204V + L180M	0.055		B2	0.020	
rtM204I + S202G + M250V	0.061		C1	0.015	
WT, GtD	0.040		C2	0.009	
			D	0.040	

- No cross-resistance with Nuc^R variants. Consistent with their distinct mechanism of action.
- AB-506 shows activity against the most prevalent HBV genotypes globally.

Table 3: Antiviral selectivity of AB-506:

Virus	Family	Genome	AB-506		Host Cell Line
			EC ₅₀ (μM)	CC ₅₀ (μM)	
HCV	Flaviviridae	(+) ssRNA	>30	>30	Huh7
WNV	Flaviviridae	(+) ssRNA	>30	>30	VERO
RSV	Paramyxoviridae	non-segmented (-) ssRNA	>30	>30	HEp2
IFA	Orthomyxoviridae	segmented (-) ssRNA	>30	>30	MDCK
HIV	Retroviridae	ssRNA to DNA	>30	>30	CEMSS
HSV1&2	Herpesviridae	dsDNA	>30	>30	VERO
HCMV	Herpesviridae	dsDNA	>30	>30	MRC5
DENV	Flaviviridae	(+) ssRNA	>22	22	BHK21
HRV	Picornaviridae	(+) ssRNA	>30	>30	H1/HeLa

- No significant inhibition of a panel of RNA & DNA viruses demonstrating high selectivity for HBV.

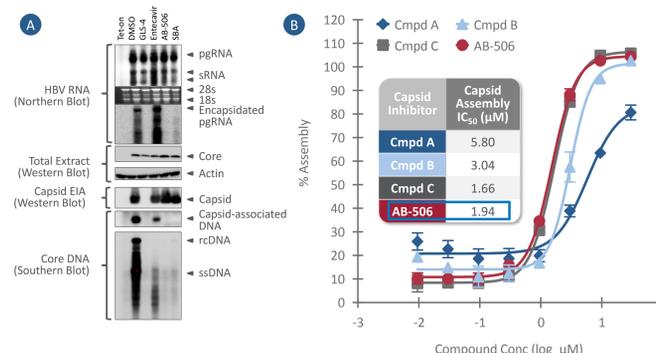


Figure 2: A) AB-506 forms empty capsids devoid of pgRNA or rcDNA in HepAD38 cells; B) Next generation capsid inhibitors accelerate capsid assembly reaction *in vitro*.

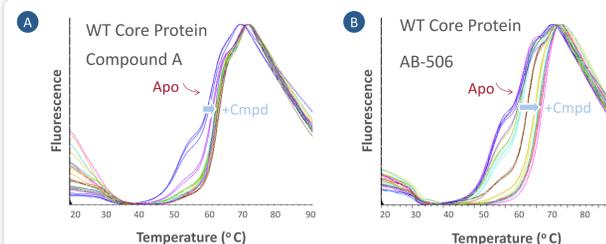


Figure 3: Increased thermal stabilization of HBV core protein by next generation capsid inhibitors. A) Compound A and B) AB-506 binding increases thermal stability of WT core protein by up to 2°C and 6°C, respectively.

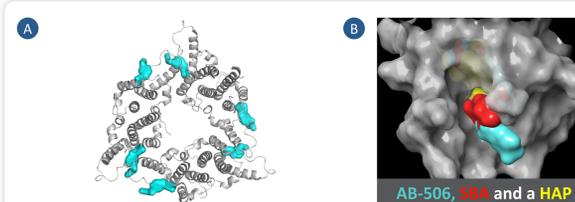


Figure 4: X-ray crystallography studies. A) AB-506 binds to core protein at the dimer:dimer interface similar to other known Class I and Class II capsid inhibitors. B) X-ray structure overlay of AB-506, a SBA and a HAP.

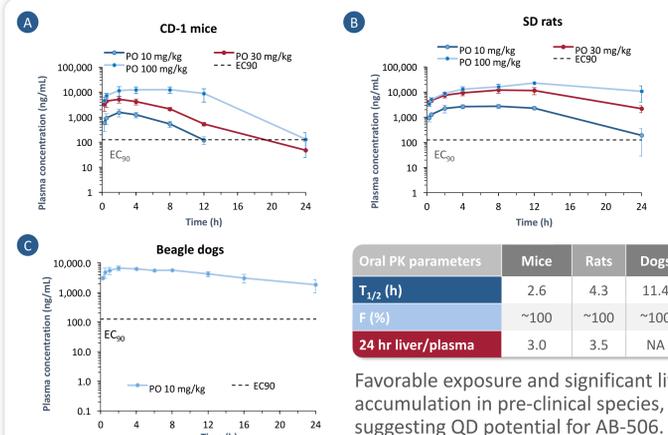


Figure 5: Pharmacokinetics of AB-506 in A) mice, B) rat, and C) dog.

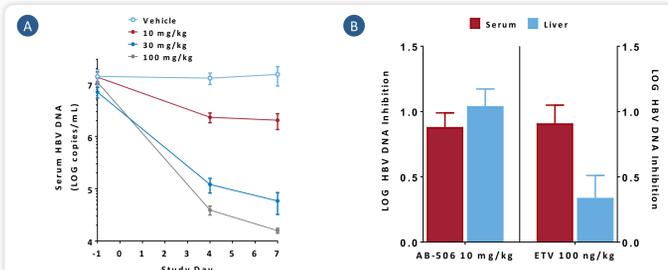


Figure 6: *In vivo* antiviral activity of AB-506. A) Reduction in serum HBV DNA is dose responsive following AB-506 administration. B) AB-506 surpassed ETV at inhibiting liver HBV DNA, at dosages where the serum HBV DNA inhibition was equivalent (data relative to vehicle at Day 7)

CONCLUSIONS

- AB-506 is a next generation highly selective HBV capsid inhibitor
- In vitro* AB-506:
 - showed potent inhibition of HBV replication in cell culture models
 - bound at the dimer:dimer interface of core protein in X-ray crystallography studies
 - inhibited pgRNA encapsidation in HepAD38 cells
 - accelerated rate of capsid assembly in a biochemical assay
 - conferred increased thermal stability to core protein indicating improved target engagement compared to first generation capsid inhibitors
- Dosing performed in multiple species suggest QD potential and significant liver concentrations achieved
- AB-506 showed potent *in vivo* activity in a HDI mouse model of HBV
- AB-506 is being evaluated for advancement into clinical development

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